Journal of Scientific Research & Reports

Journal of Kosearch Scientific Rosearch and Roports

28(10): 1-10, 2022; Article no.JSRR.90143 ISSN: 2320-0227

# Length-Weight Relationship and the Distribution of Intestinal Helminth Parasites in Freshwater Fishes from Amansea and Ebenebe Rivers in Anambra State, Nigeria

A. Okeke Obiageli <sup>a\*</sup>, Obinabo W. Obinna <sup>a</sup>, C. A. Imakwu <sup>b</sup>, Okeke C. Janefrances <sup>c</sup>, Okafor N. Chinwendu <sup>d</sup>, C. C. Nwadike <sup>a</sup>, I. O. Nnatuanya <sup>a</sup> and P. I. Afoemezie <sup>d</sup>

<sup>a</sup> Parasitology Unit, Zoology Department, Nnamdi Azikiwe University, Awka, Anambra State, Nigeria.
<sup>b</sup> Department of Parasitology and Entomology, Nnamdi Azikiwe University, Awka, Anambra State, Nigeria.

<sup>c</sup> Fisheries and Hydrobiology, Zoology Department, Nnamdi Azikiwe University, Awka, Anambra State, Nigeria.

<sup>d</sup> Technology and laboratory Unit, Zoology Department, Nnamdi Azikiwe University, Awka, Anambra State, Nigeria.

### Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

#### Article Information

DOI: 10.9734/JSRR/2022/v28i1030550

**Open Peer Review History:** 

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/90143

> Received 29 June 2022 Accepted 01 August 2022 Published 13 August 2022

Original Research Article

## ABSTRACT

**Aims:** The current study was carried out to evaluate the prevalence and distribution of intestinal helminths in freshwater fishes.

**Study Design:** The study was a survey done to establish the type of intestinal parasite found in freshwater fish in the rivers.

**Place and Duration of Study:** The fish samples were collected from Amansea and Ebenebe Rivers from November 2021 to January 2022.

Obiageli et al.; JSRR, 28(10): 1-10, 2022; Article no.JSRR.90143

**Methodology:** Intestinal parasites of three (3) fish species commonly found in these rivers (*Parachanna obscura, Clarias gariepinus, and Ctenopoma kingsleyae*) were examined. A comparative evaluation of the prevalence and distribution of the parasite was done in relation to their gender, location, weight, and length.

Results: A total of 24 fish samples were collected in the Amansea River and 26 samples were collected in Ebenebe River. Seventeen (17) samples of P. obscura, 21 samples of C. gariepinus and 12 samples of C. kingsleyae caught with the cast nets of various mesh sizes and gill traps by artisanal fishermen in Rivers were bought early in the morning, transported to the Laboratory and identified based on morphological features. Out of a total of 50 fish sampled, 17 fishes (34% of the total sample) were infected by intestinal helminths. P. obscura had the highest parasite prevalence (47.06%) followed by C. gariepinus (42.86%). However, no parasite was recorded in C. kingsleyae. Prevalence of intestinal parasites in Ebenebe (18.00%) was higher than in Amansea (16.00) with a significant difference in prevalence (p<0.05), while parasite abundance was higher in Amansea (50.94%) than in Ebenebe (49.06%) with significant difference (p<0.05). Females had the highest prevalence (35.00%) against male fishes (33.33%) with no significant difference. A total number of 53 parasites belonging to 7 genera were recovered in this study. Parasites identified were nematodes: Procamallanus sp. (41.5%), Camallanus sp. (9.43%), Rhabdochona sp. (26.42%), Contracaecum sp. (7.55%), and Spinitectus sp. (5.66%); cestodes: Polyonchobothrium sp. (5.66%); and trematodes: Clinostomum sp (3.77%). The highest parasite abundance was recorded in Procamallanus sp. (41.5%), while Clinostomum sp had the least (3.77%).

**Conclusion:** This study revealed that the prevalence of intestinal parasites in freshwater fish is relatively high. Therefore, fish consumers should subject the fish to proper processing before consumption as some of these parasites are of zoonotic importance.

Keywords: Parasites; freshwater fish; Ebenebe; Amansea; Nigeria.

## 1. INTRODUCTION

Freshwater is essential to human life. Freshwater bodies are known for their diversity of aquatic foods, their potential to contribute to sustainable healthy diets and their potential as a solution to address the "triple burden of malnutrition" (micronutrient deficiencies, undernutrition, overweight and obesity) [1]. In Nigeria, fish contributed the most essential proteins [2]. As the human population inevitably increases. the demand for fish as a source of protein also grows. In recent times, there has been a tremendous increase in the development of fish farming and culture, which is attributed to the increased need for affordable animal protein, especially in the tropics [3]. Fishes are a rich source of protein for man and other animals. They contain lipids, mineral oils, and vitamins, which all have a remarkable impact on the lives of many individuals and communities. Fish oils, which are known to aid the proper functioning of the heart, brain and immune system, contain omega-3 essential fatty acids [4]. Fish are also good sources of vitamins B6 and B12. It is also a good source of fluorine and iodine, which are needed for the development of strong teeth and the prevention of goitre [5].

Fish harvesting, handling, processing, and distribution provide a livelihood for millions of

people as well as provide foreign exchange earnings for many countries [6]. Domestic fish production in Nigeria takes place in natural water bodies such as rivers, lakes, coastal water, and seas; and artificial fish farming (pond rearing of fish in many urban areas). During the period 2007-2011, the average annual domestic fish production was put at 0.77 million metric tonnes [7].

Like other businesses, domestic fish production in Nigeria is faced with challenges, and one of them is the effects of parasitic infection on the growth and consumption of freshwater fish. These freshwater organisms harbour parasites, either external or internal, which may affect their growth. Parasitic infection constitutes a major threat to the well-being and productivity of fishery industries in Nigeria. The effects of parasitism ranged from the infliction of injuries on organs to a reduction in the population of fish, impairment of proper organ functions, and disturbance in the physiology of the fish. Its effects also include reduction of both biomass and weight through parasites feeding on the fish host [8].

The prevalence and spread of freshwater helminthes have been reported in many Rivers across Nigeria. Nematodes, acanthocephalans, trematodes and cestodes were recovered in decreasing order of abundance in a study carried out in River Niger in Illushi community. Edo State, Nigeria, Proteocephalus sp. was found in Ctenopoma kingsleve and Tilapia galilaeus. Diphyllobothrium sp. was only found in the stomach and gills of Chrysichthys nigrodigitatus. Paramphistomum sp. Trematode was found in Bucephalus sp. was found in the gills, and Synodontis eupterus Distichodus Acanthocephalans engycephalus. Also, the Pomporhynchus, Quadrigidae and Neoechinorhynchus were found on the intestine of Lates niloticus [9]. A study carried out on freshwater fish from Osse River, Benin, revealed an overall 17.1% infection rate. Of the twelve species of fish studied, Barbus sp. had the heaviest infection rate (2.2%) while Xenomystus nigrihad the least (1.0%). Parasites recovered were the tematodes Clinostomum tilapiae, Allocreadium Clinostomum sp, sp. and Diplostomum tragenna; the cestodes Polyonchobothrium sp. and Proteocephalus sp; the acanthocephalan Acanthogyrus (Acanthosentis) tilapiae: and the nematodes Camallanus sp, Procamallnus laeviconchus. Spinitectus Spirocamallanus sp, sp, Serradactnitis sp, and Spironoura sp. Nematode infection had the highest prevalence of 7.7% while trematodes recorded the least of (1.9%) [10]. Furthermore, in freshwater fish from Niger Delta tidal creek, Buguma Creek, Nigeria, only nematode parasites were encountered in the study. Of the 1,149 fish specimens examined, 213 (representing 18.5%) were infected with various nematodes parasites such as Dasyatis margarita (66.7%), followed by Pseudotolithus (Pseudotolithus) senegalensis (41.7%), while the least infected were Arius gigas and Pomadasys jubelini (3.8% and 1.4%, respectively) [11].

A review of fish species and their parasites in Nigerian water bodies showed that many parasites have being encountered by various researchers and they included protozoans such as Ichtoyobodo necator Eimeria chrysichthyic, Hexaimita sp, Tricodina sp, Crytobia ubilans, and Trypanosoma. Nematodes worms such as Cammalanus, sp Spironoura sp, Procamalus laevionchus, Gnathostoma sp, Paracamallanus, Eustrongyloides were recovered, trematodes encountered were Clinistomidae, Clinostomium tilapiae, Paramphistomium sp, Euclinustomium heterostomium, Allocreadium ghaensis, Sandonia sadanensis, and Polyoncobothrium clariae. Cestodes recovered were Proteocephalus Diphyllobothrium, sp, Amonotaenia sp, Polyoncobothrion clariae., Monobothriode wordlandi, and Heterophyid fluke.

Acanthocephalans recorded in the fishes included Neochinorhyncus rutili. Pomporhynchus Quadriaidae. Some fish hosts inhabiting parasites in various investigations across the country included Clarias gariepinus, Clarias Tilapia lazera. Synodontis sorex. zilli. niloticus, Pachynema, Oreochromis Clarias Malapterurus electricus Heterobranchus brisordalis amongst other numerous fish species [12].

Fish parasites are also medically important [13]. Parasites of *Clarias gariepinus* are of medical importance, and this raises public health concerns in the different geopolitical zones in Nigeria. This brings to question the importance of food safety in the country. Zoonotic parasites infecting fishery products are one of the several concerns in food safety. Wild fish populations are affected by parasitic diseases that directly and indirectly affect their growth and survival, hence a decline in production [14].

Researchers have shown that the list of potential fish-borne parasitic zoonoses is quite large. However, in the review, done by [15] emphasis was placed on liver fluke diseases such as clonorchiasis, opisthorchiasis and metorchiasis, as well as on intestinal trematodiasis (the heterophyids and echinostomes), anisakiasis (due to Anisakis simplex larvae). and diphyllobothriasis. Parasitic nematodes from the Anisakidae family, mostly belonging to the genus Anisakis, Pseudoterranova, Hysterothylacium, and Contracaecum cause a fish-borne zoonosis known as anisakidosis [16]. Fish-borne zoonotic infestations in humans result from ingestion of contaminated edible tissues or products of aquaculture or, to a lesser extent, from physical contact with contaminated production. Globally, over 50 species of helminth parasites from fishes and other freshwater animals are known to produce human infections with serious health hazards [17].

# 2. MATERIALS AND METHODS

# 2.1 Study Area

The study was carried out in the Ezu River of Amansea and Ebenebe Town, Awka-North Local Government Area (LGA), Anambra State, Southeastern part of Nigeria. The river runs through Amansea and Ebenebe towns. Amansea lies between latitude 6o21'40" N and longitude 6o51'38" E while Ebenebe lies between 6o20'02" N and 7o07'45" E. Ebenebe is 25km from Awka, the capital city of Anambra State and is bordered by Amansea in the south, Odoli River and Mgbakwu in the north. These areas have typical semi-tropical rainforest vegetation, characterized by freshwater swamps. They have a humid climate with a temperature of about 30.60<sup>c</sup> (870<sup>F</sup>) and a rainfall between 152 and 203cm annually [18].

### 2.2 Fish Sample Collection and Identification

A weekly collection of samples was done for a period of eight (8) weeks with the help of artisanal fishermen living in the study area that specializes in the use of fishing traps, cast and gill nets of various sizes. The fishes were collected in batches in a plastic container containing water from the river and transported to Zoology Laboratory, Nnamdi Azikiwe University Awka for analysis. The fish samples were identified with the aid of a qualified ichthyologist using the dichotomous identification method of fish determination adopted by [19]. This was achieved by taking account of the meristic features of the various fish species such as dorsal, anal, caudal, pectoral and ventral fin rays. Spines present on sampled fish were also counted and measured to aid the identification process. The morphometric data of each individual fish was taken which included the total length of the fish, head length, body depth and body girth. Fish morphometric measurements (measuring of weight and length) prior to observation of external fish condition and dissection were done. The standard lengths were measured and recorded to the nearest 0.5 centimetres (cm) using a transparent ruler on a measuring board. The standard length of each fish was taken from the tip of the snout to the end of the caudal peduncle. The weight of each fish was taken using a sensitive electronic weighing balance and recorded appropriately [20]. The total number of fish caught from the rivers was recorded; this also enabled the determination of the relative abundance of the various species in the rivers.

## 2.3 Sex Determination

The sex of each fish was determined by physical observation of the urogenital papillae which are long or distended in males while it is round and reddish in matured females. The sex was also confirmed by internal examination after dissection to expose the paired testes in the males and paired ovaries in the females [21].

## 2.4 Sample Analysis

The ventral surface of each sample was cut open lengthwise to expose the alimentary canal. The stomach and intestine of each of the fish were cut open, and the contents were washed into Petri-dish containing normal saline. The lining of the gut lumen was properly washed in normal saline with macroscopic helminths picked with a soft camel hair brush and thereafter, the remaining contents were separated by sedimentation through centrifugation using Techmel Centrifuge 800D and then decanted. One to two drops of the preparation were placed on a microscopic slide covered with slip and observed under an x4 and x10 binocular light microscope for various intestinal parasites. The parasites were identified with the help of a qualified parasitologist according to the methods [22]. Fish specimens found with the parasite were given separate serial numbers to differentiate them from those without parasites. Parasites obtained were counted and labelled with the serial number of the fish. The recovered parasites were fixed in heated 70% ethanol, counted, and recorded. The hot fixative caused them to straighten out making them easier for further studies and examination.

## **2.5 Statistical Analysis**

The results were analyzed descriptively to calculate the prevalence and distribution of the parasite. The abundance of intestinal parasites and the prevalence of intestinal parasites in the samples were calculated using Microsoft Excel. The chi-square test ( $\chi$ 2) was used to determine the differences in parasitaemia amongst the fish species from the two locations. The level of significant difference was set at P<0.05.

# 3. RESULTS

Table 1 revealed that *Procamallanus sp.* (68.2%), *Contracecum sp.* (100%), *Camallanus sp.* (80.00%), *Rhabdochona sp.* (71.4%), and *Spinitectus sp.* (100.0%) were more abundant in the fish *P. obscura* in comparison to *Clinostomum sp.* (100.0%) and *Polyonchobothrium sp.* (100.0%) was found in *C. gariepinus.* There was a significant difference in the abundance of parasite species compared to the fish species sampled (P<0.05).

Intestinal parasites of fish species were more abundant in Amansea (50.94%) than in the Ebenebe River (49.06%). *Procamallanus sp.* was more abundant (41.51%) followed by *Rhabdochona sp.* (26.42%) while *Clinostomum sp.* had the least (3.77%). There was a significant difference in the number of parasite species recovered between locations (P<0.05) (Table 2).

The result in Table 3 revealed that out of the 50 fish sampled and sacrificed, 17(34.00%) were infected with intestinal parasites. The females had a higher prevalence (35.00%) than the male fishes (33.33%). There was no significant difference in the prevalence of intestinal parasites in fish sampled in relation to sex (P>0.05).

*P. obscura* had the highest parasite prevalence, 8(47.06%) when compared to *C. gariepinus*,

9(42.86%) and *C. kingsleyae*, 0(0.00%). Also, the prevalence of intestinal parasites in Ebenebe, 9(18.00%) was higher than in Amansea 8(16.00%). However, there was a significant difference in the prevalence of intestinal parasites in relation to fish species sampled (P<0.05) (Table4).

The study revealed that fish weighing 400-499.9g had the highest prevalence of intestinal parasites, 3(100.00%) than those weighing 200-299.9g, 6(35.29%) and 100-199.9g 7(28.00%) while those weighing 300-399.9g, had the least, 0(0.00%). There was a significant difference in the prevalence of intestinal parasites in relation to the weights of fishes sampled (P<0.05) (Table 5).

#### Table 1. Parasites distribution by species of fish sampled

C. Total yae gariepinus
b) 7(31.8%) 22(41.51%)
b) 0(0.00%) 4(7.55%)
) 1(20.0%) 5(9.43%)
b) 4(28.6%) 14(26.42%)
b) 0(0.00%) 3(5.66)
b) 2(100.0%) 2(3.77%)
) 3 (100.0%) 3 (5.66)
b) <b>17(</b> 32.1%) <b>53(100)</b>
2

#### Table 2. Abundance of parasites in relation to location

Abundance of p	Total (%)	
Amansea	Ebenebe	
16(59.26)	6(23.08)	22(41.51)
4(14.81)	1(3.85)	5(9.43)
3(11.11)	11(42.31)	14(26.42)
4(14.81)	0(0.00)	4(7.55)
0(0.00)	3(11.54)	3(5.66)
0(0.00)	2(7.69)	2(3.77)
0(0.00)	3(11.54)	3(5.66)
27 (50.94)	26(49.06)	53(100)
	Amansea           16(59.26)           4(14.81)           3(11.11)           4(14.81)           0(0.00)           0(0.00)           0(0.00)           0(0.00)	$\begin{array}{cccc} 16(59.26) & 6(23.08) \\ 4(14.81) & 1(3.85) \\ 3(11.11) & 11(42.31) \\ 4(14.81) & 0(0.00) \\ 0(0.00) & 3(11.54) \\ 0(0.00) & 2(7.69) \\ 0(0.00) & 3(11.54) \end{array}$

 $\chi^2 = 22.906$ , df =6, P =0.001

#### Table 3. Prevalence of intestinal parasites in relation to the sex of fish

Sexes	No. Examined	No. Infected	Prevalence	
Male	30	10	33.33	
Female	20	7	35.00	
Total	50	17	34.00	

		Amansea	Ebenebe	
Fish species	No. Examined	No. Infected	No. Infected	Total
P. obscura	17	6(35.29)	2(11.76)	8(47.06)
C. gariepinus	21	2(9.52)	7(33.33)	9(42.86)
C. kingsleyae	12	0(0.00)	0(0.00)	0(0.00)
Total	50	8(16.00)	9(18.00)	17(34.00)

# Table 4. Prevalence of intestinal parasites in relation to fish species sampled from the two Rivers

Weight range	No.	Number of fishes infected (%)			
(g)	Examined	P. obscura	C. gariepinus	C. kingsleyae	Total
100-199.9	25	4(16.00)	4(16.00)	0(0.00)	8(32.00)
200-299.9	17	1(5.88)	5(29.41)	0(0.00)	6(35.29)
300-399.9	5	0(0.00)	0(0.00)	0(0.00)	0(0.00)
400-499.9	3	3(100.00)	0(0.00)	0(0.00)	3(100.00)
Total	50	7(16.00)	9(18.00)	0(0.00)	17(34.00)

Table 6. Prevalence of intestinal	parasites in relation to lengths
Table 0. Trevalence of intestinal	

Fish species				
Number	P. obscura	C. gariepinus	C. kingsleyae	
examined	No. Infected (%)	No. Infected (%)	No. Infected (%)	-
12	2(16.67)	3(25.00)	0(0.00)	5(16.67)
32	3(9.38)	5(15.63)	0(0.00)	8(28.13)
6	3(50.00)	1(16.67)	0(0.00)	4(66.67)
50	8(16.00)	9(18.00)	0(0.00)	17(34.00)
	examined 12 32 6	Number examined         P. obscura           No. Infected (%)           12         2(16.67)           32         3(9.38)           6         3(50.00)	Number examined         P. obscura No. Infected (%)         C. gariepinus           12         2(16.67)         3(25.00)           32         3(9.38)         5(15.63)           6         3(50.00)         1(16.67)	Number examined         P. obscura         C. gariepinus         C. kingsleyae           No. Infected (%)         No. Infected (%)         No. Infected (%)         No. Infected (%)           12         2(16.67)         3(25.00)         0(0.00)           32         3(9.38)         5(15.63)         0(0.00)           6         3(50.00)         1(16.67)         0(0.00)

Table 6 revealed that fish with lengths ranging from 30-39.9 cm had the highest prevalence of parasites, 4(66.67%) than those from 20-29.9 cm, 9(28.13%) while those with lengths between 10-19.9 cmhad the least, 5(16.67%) infection. There was no significant difference in the prevalence of intestinal parasites in relation to the length of fish sampled.

## 4. DISCUSSION

A total of fifty-three (53) helminths belonging to seven (7) helminth genera were recovered in this study, which includes trematodes: *Clinostomum* sp., cestodes: *Polyonchobothrium* sp., and nematodes: *Procamallanus* sp., *Camallanus* sp., *Contracaecum* sp., *Rhabdochona* sp., and *Spinitectus* sp. The recovery of similar helminth parasites has been reported previously from the same or related species elsewhere [23, 24]. In the Ebonyi River in Ebonyi State, Nigeria, the four freshwater fishes studied harboured helminth parasites [25]. The high prevalence of helminth parasites in freshwater fish may be attributed to anthropogenic activities by humans who harbour the parasites' eggs. Secondly, freshwater fish tend to have more parasites (e. g. in the United States) compared to their oceanic counterparts [26] due to factors such as ecological and biological factors that shape the pattern and prevalence of infection amongst fish species and location. These factors are summarised as human activity and natural factors [27].

The prevalence of helminth parasites was higher in the Ebenebe community compared to Amansea, with a significant difference (p 0.05). Freshwater (FW) fish inhabit aquatic environments that vary greatly in their ionic composition and pH, but uniformly these environments have much lower concentrations of total salts than the fish's body fluids [28]. Furthermore, the type of habitat, water pollution, over-exploitation, the spread of invasive species, the spread of alien parasites and pathogens, salinization, acidification, and climate change may play vital roles in the prevalence differences [29].

This study also showed a high parasitic infection in P. obscura, followed by C. gariepinus, while no parasite was recovered from C. kingsleyae. In contrast to the findings of this study, freshwater fishes from the River Niger Proteocephalus species were recovered in C. kingslevae [30].Forty-one per cent (41.67%) of infection in 120 examined C. gariepinus samples has been reported elsewhere [24], which when compared to this study is of lower prevalence, although a smaller number of samples were used in this study. Also, in another study conducted in Iran, 18.96% of the examined catfish were infected with digenean trematodes (39%) and cestodes (34%) [31]. In Pakistan, a total of 43 specimens of four species of freshwater fish gut contents and visceral organs revealed the highest prevalence of nematodes (60.46%), followed by trematodes (53.48%). The differences in the prevalence of parasites in P. obscura and C. kingsleyae may be attributed to a number of factors which may include preferred location in the habitat, immuno-competence of the fish, the behavioural and feeding pattern of the fish, as well as the high nutritional content of their intestines and suitability of the fish host in the provision of appropriate ecological requirements of the parasite [32].

Knowledge of the level of parasitisation of fish in a particular population is important because the damage associated with it is relative to the rate of infestation with the parasite. Heavily infected fish may have mechanical damage such as fusion of gill lamellae, tissue replacement, and physiological damage such as cell proliferation, immune-modulation, detrimental behavioural responses, altered growth, and reproductive damage. Changes in the environment, both anthropogenic and environmental, can alter the parasite/host equilibrium and cause disease or mortality in fish [33].

This study also shows that males have a slightly lower prevalence rate than female fish. There was no significant difference in prevalence in relation to gender. A higher prevalence in females (42.86%) compared to their male counterparts (40.00%) was reported in some other studies [24, 30]. It has been reported that infection of the four fish species in the Ebonyi River was not significantly sex-dependent [33]. Variations in parasitic infection among the sexes of fish may be by chance and not for any other reason [34]. The gender difference recorded in this study for the helminth-infected fishes could be due to alterations in the physiological status of sampled fishes, particularly males, which could be a result of differential feeding either in quality or quantity of food eaten. The male may have a more powerful immune system than his female counterpart. It could also mean that more females were more readily available for parasitic infestation than males.

There is a relationship between parasitic prevalence infections and fish length and weight, which are equivalent to fish age. Fish weighing 400-499.9g and measuring 30-39.9cm in length had a higher prevalence of parasitic infection than fish of lower weights and lengths. The relationship could be explained by differences in their feeding habits caused by age, as older fishes are more likely to pick up parasites than younger fishes because they supposedly feed more [35]. Another reason could be that younger fish, which are thought to be more active and have better immune systems, were less likely to get parasites than older fish, which had weaker immune systems.

This study also reported the highest abundance of nematodes (*Procamallanus sp., Rhabdochona* sp., *Camallanus* sp., *Contracaecum* sp., and *Spinitectus* sp.), followed by a corresponding low abundance of cestodes (*Polyonchoboyhrium* sp.) and trematodes (*Clinostomum* sp.). Similar helminth parasites were recovered in similar studies [23]. However, *Polyonchobothrium* sp. and *Procamallanus* sp. recovered in this study was not among the parasite fauna. However, cestode (*Polyonchobothrium clarias*, 28.18%) and nematode (*Procamallanus laevionchus*, 11.82%) [24] have also been recovered in a similar study.

Contracaecum sp. is among the intestinal parasite recovered from the fish samples that is of zoonotic importance. It is one of the gastrointestinal parasites of marine mammals fish-eating birds. Small, shrimplike and crustaceans (krill) serve as the first intermediate host and the infective stages (L3 larvae) are found in many saltwater fish and squid paratenic hosts and these are possible sources for human infection [36]. In a study of one thousand, one hundred and fifteen (1,115) fish specimens belonging to nine fish species collected from Lake Nasser, Egypt, four (4) fish species (Oreochromis niloticus, Tilapia galilaea, Lates *niloticus*, and *Hydrocynus forskahlii*) were found to be infected with third stage larvae of *Contracaecum* sp. only [37].

Treatment is with surgical or endoscopic removal, appropriate education of the public, maintenance of good hygiene, adoption of good self/personal-hygiene, the practice of eating properly cooked food, being cautious of what to eat when travelling to developing countries and the use of clean water for drinking and bathing. All or some of these prevention and control practise can abate the worldwide fish-borne zoonoses [38].

# 5. CONCLUSION

The study revealed that the helminth parasites distribution was higher in females with no significant difference in prevalence. The study also reported a higher prevalence of intestinal parasites in Ebenebe than in Amansea with a recorded significant difference. Seven parasite species, namely, *Procamallanus sp.* (nematode), *Camallanus sp.* (nematode), *Rhabdochona sp.* (nematode), *Contracaecum sp.* (nematode), *Polyonchobothrium sp.* (cestode), *Clinostomum sp.* (trematode), and *Spinitectus sp.* were the intestinal parasite found in 3 fish species from Amansea and Ebenebe Rivers.

# 6. RECOMMENDATION

It is necessary to always carry out surveys to determine and identify the parasite fauna and their distribution in different fish species in freshwater habitats such as Amansea and Ebenebe Rivers in Anambra State. Further studies should be encouraged to help establish the relationship between the physicochemical parameters and parasites assemblage. Also effective control measures against fish-borne zoonotic parasites should take note of the numerous factors accountable for transmission and occurrence.

# ETHICAL APPROVAL

This study followed guidelines for the care and use of experimental animals established by the Animal Care and Use Committee of Nnamdi Azikiwe University, Awka, Nigeria, for the control and supervision of experiments on animals (RE: NAU/AREC/2021/00046).

# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

## REFERENCES

- 1. United Nations, FINAL-UN-Nutrition-Aquatic-foods-Paper\_EN .pdf. The role of aquatic foods in sustainable healthy diets. UN Nutrition Discussion Paper; 2021.
- De Vries-Ten Have J, Owolabi A, Steijns J, Kudla U, Melse-Boonstra A. Protein intake adequacy among Nigerian infants, children, adolescents and women and protein quality of commonly consumed foods. Nutrition Research Reviews. 2020;33(1):102–120. Available:https://doi.org/10.1017/S0954422 419000222
- 3. Adegbesan SI, Obasa SO, Abdulraheem I. Growth performance, haematology and histopathology of African catfish (*Clarias gariepinus*) fed varying levels of Aloe barbadensis leaves. J Fish. 2018;6(1):553-62.
- 4. Balami S, Sharma A, Karn R. Significance of nutritional value of fish for human health. Malays J Halal Res. 2019;2(2):32-4 [online].
- Ahmed M, Liaquat M, Shah AS, Abdel-Farid IB, Jahangir M. Proximate composition and fatty acid profiles of selected fish species from Pakistan. J Anim Plant Sci. 2020;30:869-75.
- Adeyeye SAO, Oyewole OB. An overview of traditional fish smoking in Africa. J Culinary Sci Technol. 2016;14(3):198-215.
- Udoh EJ, Akpan SB. Macroeconomic variables affecting fish production in Nigeria. Asian J Agric Rural Dev. 2019; 9(2):216-30.
- 8. Adebambo AA. Fish species parasites: a review in Nigerian water bodies. J Res For Wildl Environ. 2020;12(3).
- 9. Onyedineke NE, Obi U, Ofoegbu PU, Ukogo I. Helminth parasites of some freshwater fish from River Niger at Illushi, Edo State, Nigeria. J Am Sci. 2010;6:3.
- 10. Okaka CE, Akhigbe JE. Helminth parasites of some tropical freshwater fish from Osse River in Benin, southern Nigeria. The J Freshw Biol. 2004;8(1).
- 11. Ogbeibu AE, Okaka CE, Oribhabor BJ. Gastrointestinal helminth parasites community of fish species in a Niger Delta tidal Creek, Nigeria. J Ecosyst. 2014;2014:1-10.
- 12. Adebambo AAR. Fish species parasites: a review in Nigerian water bodies. J Res For Wildl Environ. 2020;12(3):2141-1778.

- Odoh VU, Abuh OO, Haruna MM, Yisa MA, Bids AA. Medically Important Parasites of Clarias garipienus (catfish) in Nigeria. Adv Biotechnol Microbiol. 2019;15(1):555904.
- 14. Quiazon KMA. Updates on aquatic parasites in fisheries: implications to food safety, food security and environmental protection. J Coast Zone Manag. 2015; 18(1).
- Chai JY, Darwin Murrell K, Lymbery AJ. Fish-borne parasitic zoonoses: status and issues. Int J Parasitol. 2005;35(11-12):1233-54.
- Farinha AP, Moreira M, de Magalhães CP, Schrama D, Cerqueira M, Carrilho R et al. Proteomics for quality and safety in fishery products. Galanakis CM, editor. Sustainable fish production and processing. Academic Press. 2022; 45-78. Available:https://www.sciencedirect.com/sc ience/article/pii/B9780128242964000074.
- 17. Avishek B. Fish-borne parasites proficient in zoonotic diseases: a mini review. Insights Vet Sci. 2022;6(1):005-12.
- Onyido AE, Zeibe CC, Okonkwo NJ, Ezugbo-Nwobi IK, Egbuche CM, Udemezue IO et al. Damage caused by the bean Bruchid, Callosobruchus maculatus (Fabricius) on different legume seeds on sale in Awka and Onitsha Markets, Anambra State, South Eastern Nigeria. Afr Res Rev. 2011;5(4):116-23.
- Odo GE, Didigwu NC, Eyo JE. The fish fauna of Anambra river basin, Nigeria: species abundance and morphometry. Rev biol trop. 2009;57(1-2):177-86.
- 20. Bagbe AS. Statistical Evaluation of the Link Between intestinal Helminth Parasites and Sex, Weight and Length of Clarias gariepinus in a Coastal Community in Okitipupa Local Government Area, Ondo State. Int J Prog Sci Technol. 2021;30(1):73-83.
- 21. Imam TS, Dewu RA. Survey of Piscine ecto and intestinal parasites of Clarias sp. sold at Galadima road fish market. Kano: metropolis, Nigeria. Bioscience Research Communication. 2010;22(4):209-14.
- 22. Pugachev ON, Gerasev PI, Gussev AV, Ergens R, Khotenowsky. Guide to Monogenoidea of Freshwater fish of Palaearctic and Amur regions. Milano, Italy: LedizioniLedi publishing; 2010.
- 23. Onoja-Abutu AE, Okpanachi MA, Alkazmi L, Yaro CA, Batiha GES. Branchial chamber and intestinal tracts parasites of

fish species in Benue and Niger Rivers, North Central, Nigeria. Int J Zool. 2021;2021:10.

- Idris HS, Balarabe-Musa B, Osawe SO. The incidence of endo-parasites of Clarias gariepinus (Sharp tooth catfish) (Burchell, 1822) and Oreochromis niloticus (tilapia fish) (Linnaeus, 1958) in Jeremiah Usein River, Gwagwalada, Nigeria. Intercontinental J Biol Sci. 2013;1(1):1-5.
- 25. Onyishi GC, Aguzie ION. Survey of helminth parasites of fish in Ebonyi River at Ehaamufu, Enugu State, Nigeria. Anim Res Int. 2018;15(3):3112-9.
- 26. Available:https://massivesci.com/notes/fres hwater-ocean-fish-sushi-food. Parasites in freshwater fish make them more hazardous than ocean fish.
- Kołodziej-Sobocińska M. Factors affecting the spread of parasites in populations of wild European terrestrial mammals. Mamm Res. 2019;64(3):301-18.
- Wilson RW 2011. Role of the gut | Gut Ion, Osmotic and acid-base Regulation in Encyclopedia of Fish Physiology, Freshwater Fish – an overview https://www.sciencedirect.com > fr.
- 29. Mehner T, Brucet S. Structure of Fish Communities in Lakes and Its Abiotic and Biotic Determinants in Reference Module in Earth Systems and Environmental Sciences, Freshwater Fish – an Overview; 2021.

Available: https://www.sciencedirect.com > fr.

- Onyedineke NE, Obi U, Ofoegbu PU, Ukogo I. Helminth parasites of some freshwater fish from river Niger at Illushi, Edo State, Nigeria. J Am Sci. 2010;6(3).
- Yakhchali M, Ali-Asghar T, Mozafar G. The occurrence of helminth parasites in the intestinal of catfish (Silurus glanis Linnaeus 1758) from the Zarrine-roud river, Iran. Vet Res Forum. 2012;3(2):143-5.
- 32. Ukuru MN, Adikwu IA. Seasonal prevalence of parasites of Clariids fishes from the Lower Benue River, Nigeria. Niger J Fish Aquacult. 2017;5:11-9.
- Iwanowicz DD. Overview on the effects of parasites on fish health. 11649 Leetown Road, Kearneysville, WV: United States Geological Survey, Leetown Science Center, National Fish Health Research Laboratory. p. 25430. A Review Conference paper; 2014. Available:https://www.researchgate.net/pu blication/230635404.

- Biu AA, Akorede GJ. Prevalence of endoparasitized Clarias gariepinus (Burchell 1822) in Maiduguri, Nigeria. Niger J Fish Aquacult. 2013;1(1):1-6.
- 35. Mgbemena A, Arimoro F, Omalu I, Keke U. Prevalence of helminth parasites of Clarias gariepinus and Tilapia zilli in relation to age and sex in an afro-tropical stream. Egypt J Aquat Biol Fish. 2020;24(5):1-11.
- McPherson RA: Chapter 65. Medical parasitology. Available:https://www.clinicalkey.com/#!/co ntent/book/3-s2.0-

B9780323673204000651. In: Henry's clinical diagnosis and management by laboratory methods. 2022; 1290-1351.e3.

- Younis AE, Saad AI, Rabei JM. The occurrence of Contracaecum sp. larvae (Nematoda: Anisakidae) in four teleostean species from Lake Nasser, Egypt: morphological and molecular studies. JoBAZ. 2017;78(1):9.
- Shamsi S. Parasite loss or parasite gain? Story of contracaecum nematodes in antipodean waters. Parasite Epidemiol Control. 2019;4. 2020:e00087.

© 2022 Obiageli et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/90143